

Inhibition of dimethylarginine dimethylaminohydrolase (DDAH) and arginine deiminase (ADI) by pentafluorophenyl (PFP) sulfonates†

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A range of pentafluorophenyl (PFP) sulfonate esters derived from the reaction of PFP vinyl sulfonate and various nitrones are shown to have significant inhibitory activity against the bacterial enzymes DDAH and ADI.

Nitric oxide (NO) is an important mediator of intracellular signaling and has attracted interest as a target for therapeutic intervention, as it is widely acknowledged that there are a variety of disease states for which NO is implicated.¹ One of the most significant problems associated with the design of inhibitors, is to target the pathological excess NO production without disrupting essential NO-mediated processes, often by seeking selectivity for a particular NOS isoform. One method for potentially circumventing these problems is the indirect modulation of NO levels by inhibition of the enzyme dimethylarginine dimethylaminohydrolase, DDAH which is responsible for controlling levels of *N*^G-methyl-L-arginine (MMA) and *N*^G, *N*^G-dimethyl-L-arginine (ADMA) which are endogenous inhibitors of NOS.^{2,3}

Inhibition of bacterial DDAH⁴ is also of interest as it offers opportunities for the development of new anti-bacterial agents. The structurally related enzyme arginine deiminase (ADI) is also a possible antibacterial/antiprotozoal target, as various pathogenic organisms utilize ADI to generate ATP under anaerobic conditions.^{5a}

Recently high-resolution structures of a bacterial DDAH⁶ and ADI⁵ have been disclosed and it has been shown in both enzymes that the active site comprises a catalytic triad containing an acidic residue (Glu/Asp), a basic residue (His) and a cysteine residue (Cys). Both enzymes are known to catalyze the conversion of the substrate(s) MMA and ADMA to citrulline as shown in Fig. 1.

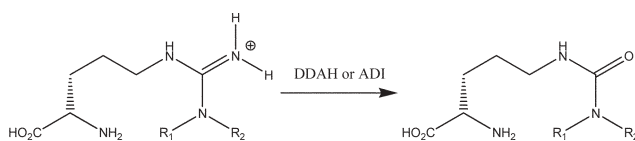


Fig. 1 Conversions of arginine ($R_1 = R_2 = H$), ADMA ($R_1 = R_2 = Me$) and/or MMA ($R_1 = Me, R_2 = H$) to citrulline are catalysed by DDAH and ADI.

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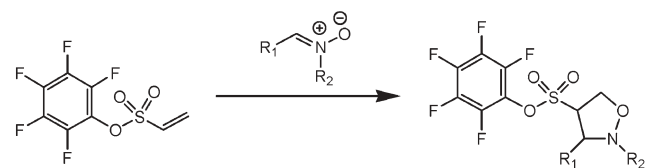
In a recent communication Knipp *et al.* described the cysteine modification of DDAH by HcyNO and proposed this as a lead for the possible development of covalent inhibitors of DDAH and ADI.⁷ In that work it was proposed that it should be possible to rationally design covalent inhibitors of DDAH based on those findings. That work has prompted us to disclose our own studies, which identify novel small molecule inhibitors of DDAH and ADI. Whilst the development of small molecule inhibitors of both DDAH and ADI is appealing, it is notable that there is only one known inhibitor of bacterial DDAH, which has modest affinity and is an arginine homologue,⁸ and there are no known inhibitors of ADI. Herein we disclose our preliminary studies on the use of pentafluorophenyl (PFP) sulfonates as an unprecedented new class of enzyme inhibitors. The biological activity of the PFP-sulfonate group is completely unexplored and is highlighted here by the development of inhibitors of DDAH and ADI.

As had previously been noted by one of us in the disclosure of the crystal structure of DDAH, the active site resembles that of a cysteine protease with a catalytic triad.⁵

The work of Roush *et al.* on the use of sulfonates and sulfonamides as inhibitors of cysteine proteases,⁹ stimulated us to speculate that it may be possible to generate non-covalent inhibitors of DDAH and ADI based on sulfonates and sulfonamides or closely related structures. This would offer an opportunity to develop molecular scaffolds, which would be markedly different in their structure to arginine mimetics, which may be a more obvious class of potential inhibitor. In order to test this speculative hypothesis we decided to evaluate a diverse collection of heterocyclic PFP-sulfonates as potential inhibitors of DDAH and ADI.

Our previously disclosed synthetic approach to such species was based on the 1,3-dipolar cycloaddition reaction of a PFP-sulfonate with nitrones (Scheme 1).¹⁰

An initial screen of a variety of PFP-sulfonates and related structures at relatively high concentrations (500 μ m, data not shown) provided some encouraging inhibition of both *pseudomonas* enzymes DDAH and ADI. At 50 μ m a smaller selection of



Scheme 1 Synthetic approach to heterocyclic PFP-sulfonates via 1,3-dipolar cycloaddition.

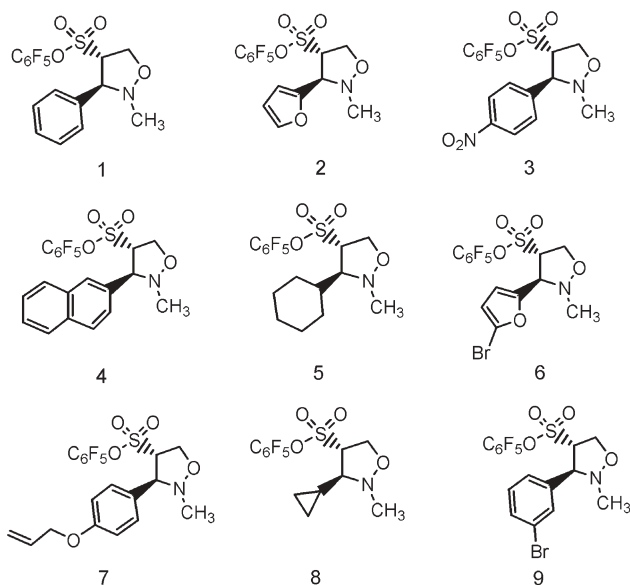


Fig. 2 Molecular structures of PFP sulfonates exhibiting significant inhibition of PaDDAH and PaADI.

Table 1 Activity of PFP-sulfonates against PaDDAH and PaADI (1 mM substrate)

| Entry | DDAH Inhibition at 50 μ M (%) | IC ₅₀ /DDAH, μ M | ADI Inhibition at 50 μ M (%) | IC ₅₀ /ADI, μ M |
|-------|-----------------------------------|---------------------------------|----------------------------------|--------------------------------|
| 1 | 30 | — | 14 | — |
| 2 | 63 | 34 | 27 | 246 |
| 3 | 76 | 21 | 35 | 74 |
| 4 | 56 | 32 | 33 | 167 |
| 5 | 40 | — | 26 | — |
| 6 | 65 | 16 | 38 | 103 |
| 7 | 44 | — | 14 | — |
| 8 | 41 | — | 15 | — |
| 9 | 58 | 58 | 27 | — |

PFP sulfonates retained significant activity and their molecular structures are shown in Fig. 2.

As can be seen from Table 1 the majority of these compounds were found to have activity against both DDAH and ADI at 50 μ M concentration. These preliminary data indicate that there is

greater inhibition of DDAH compared with ADI. IC₅₀ values were determined for a small selection of the most active species and it can be seen that compounds **2**, **3**, **4**, **6** and **9** exhibit significant activity against DDAH and compounds **3** and **6** also show significant activity against ADI.

These data suggest that there is potential for considerable optimization to give a new series of inhibitors based on the PFP-isoxazolidine structural motif.

In order to assess the nature of the inhibition we carried out experiments on reversibility and time-dependence (Fig. 3).

Inhibition by compound **6** can be at least partially reversed by addition of increased amounts of substrate (Fig. 3(a)), suggesting competitive inhibition. The large excess of substrate required to reverse inhibition may indicate that the inhibitor is more tightly bound than the substrate. In a separate experiment, compound **3** exhibited a constant level of DDAH inhibition over an 80-min period (Fig. 3(b)). Covalent inhibitors would show a time-dependent increase in inhibition as the enzyme becomes progressively irreversibly bound, therefore the data suggests that our inhibitors are not acting by a covalent mechanism. This is consistent with the observed reversibility of inhibition.¹¹

In summary we have described new inhibitors of the enzymes DDAH and ADI. From these experiments it would appear that these PFP-sulfonates may be reversible inhibitors of DDAH. Irrespective of the detailed mechanism underlying the inhibition, this is the first time that non-substrate-like inhibitors for DDAH have been identified and these are the most potent inhibitors of bacterial DDAH currently known. Moreover these results identify the first small molecules to inhibit the enzyme ADI. The present study has also demonstrated that the PFP-sulfonate motif may play an important role in future studies directed toward identification of small molecule enzyme inhibitors and/or ligands for proteins. The simplicity with which diverse arrays of PFP-derivatives can be prepared may facilitate further small-molecule discovery activities. The further development of this work to identify details of the molecular interaction of these PFP-sulfonate derivatives with DDAH and ADI is under way.

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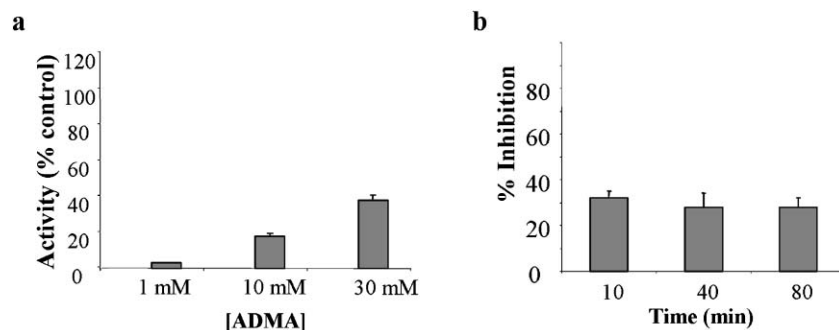


Fig. 3 (a) Effect of increasing substrate concentration on inhibition of PaDDAH by compound **6**, 75 μ M. (b) Time-dependence of PaDDAH inhibition by compound **3**, 10 μ M.

Notes and references

- 1 (a) J. M. Fukuto and G. Chaudhuri, *Annu. Rev. Pharmacol. Toxicol.*, 1995, **35**, 165–194; (b) S. Moncada and A. Higgs, *FASEB J.*, 1995, **9**, 1319–1330; (c) A. J. Hobbs, A. Higgs and S. Moncada, *Annu. Rev. Pharmacol. Toxicol.*, 1999, **39**, 191–220.
- 2 (a) P. Vallance and J. M. Leiper, *Nat. Rev. Drug Discov.*, 2002, **1**, 939–950.
- 3 T. Ogawa, M. Kimoto and K. Sasaoka, *J. Biol. Chem.*, 1989, **264**, 10205–10209.
- 4 J. Santa Maria, P. Vallance, I. G. Charles and J. M. Leiper, *Mol. Microbiol.*, 1999, **33**, 1278–1279.
- 5 (a) A. Galkin, L. Kulakova, E. Sarikaya, K. Lim, A. Howard and O. Herzberg, *J. Biol. Chem.*, 2004, **279**, 14001–14008; (b) K. Das, G. H. Butler, V. Kwiatkowski, A. D. Clark Jr., P. Yadav and E. Arnold, *Structure*, 2004, **12**, 657–667.
- 6 J. Murray-Rust, J. M. Leiper, M. McAlister, J. Phelan, S. Tilley, J. Santa Maria, P. Vallance and N. McDonald, *Nat. Struct. Biol.*, 2001, **8**, 679–683.
- 7 M. Knipp, O. Braun and M. Vašák, *J. Am. Chem. Soc.*, 2005, **127**, 2372–2373.
- 8 (a) R. J. MacAllister, H. Parry, M. Kimoto, T. Ogawa, R. J. Russell, H. Hodson, G. St. J. Whitley and G. P. Vallance, *Br. J. Pharmacol.*, 1996, **119**, 1533–1540; (b) S. Rossiter, C. L. Smith, M. Malaki, M. Nandi, H. Gill, J. M. Leiper, P. Vallance and D. L. Selwood, *J. Med. Chem.*, 2005, **48**, 4670–4678.
- 9 (a) W. R. Roush, S. L. Gwaltney, J. M. Cheng, K. A. Scheidt, J. H. McKerrow and E. Hansell, *J. Am. Chem. Soc.*, 1998, **120**, 10994–10995; (b) W. R. Roush, J. M. Cheng and B. Knapp-Red, *Bioorg. Med. Chem. Lett.*, 2001, **11**, 2759–2762; (c) B. R. Shenai, A. Alvarez-Hernandez, P. Y. Chong, C. D. Emal, R. J. Neitz, W. R. Roush and P. J. Rosenthal, *Antimicrob. Agents Chemother.*, 2003, **47**, 154–160.
- 10 S. Caddick and H. D. Bush, *Org. Lett.*, 2003, **5**, 2489–2492.
- 11 J. D. Winkler, C. M. Sung, M. Chabot-Fletcher, D. E. Griswold, D. L. A. Marshall, F. H. Chilton, W. Bondinell and R. J. Mayer, *Mol. Pharmacol.*, 1998, **53**, 322–329.



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